

Ver. 1902-00

# Punch-it™ NA-Sample Kit

#### **Kit Contents**

Punch-it™ NA-Sample Kit			
Cat. No.	PINA15 (15 preps/kit)	PINA50 (50 preps/kit)	PINA200 (200 preps/kit)
Punch-it <sup>™</sup> kit	15ea	50ea	50ea x 4ea
Puncher	15ea	50ea	50ea x 4ea
Lysis buffer	15 ml	50 ml	50 ml x 4ea
Washing buffer	1.8 ml (Add 7.2 ml ethanol)	6 ml (Add 24 ml ethanol)	6 ml x 4ea (Add 24 ml ethanol)
Instruction for Use	1ea	1ea	1ea

**X** Store at room temperature

# Description

**Punch-it<sup>™</sup> NA-Sample Kit** is a nucleic acid purification kit designed for instant isolation of total nucleic acids (DNA and RNA) from a little amount of various samples such as animals, blood, plants, bacteria and viruses. Using the paper chromatographic method, enzyme inhibitors are efficiently removed from the samples by this kit. The purified nucleic acids bound on membrane could be used directly in PCR or RT-PCR reactions as templates.

## **Applications**

Purification of nucleic acid for PCR or RT-PCR

Purification of nucleic acid for molecular diagnostics

## **Quality control**

Punch-it<sup>™</sup> NA-Sample Kit was tested for the isolation of total nucleic acids from various samples such as animals, blood, plants, bacteria and viruses. Nucleic acid purified with Punch-it<sup>™</sup> NA-Sample Kit was evaluated by the amplification of target DNA or transcript using PCR or RT-PCR.

## Important thing to do before starting

Before using Washing Buffer, add absolute ethanol according to the bottle label to obtain a working solution, You may use 80% ethanol, instead of Washing Buffer.

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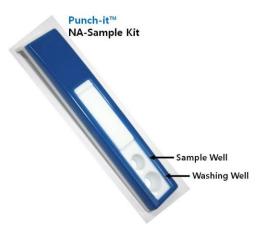


#### **Protocol**

# **Sample Preparation**

### [Whole Blood, saliva or purified virus]

- 1) Prepare **50** µl of Lysis Buffer into a 1.5 ml microcentrifuge tube (not provided).
- 2) Add 5 ~ 10 μl of sample into the tube including lysis buffer and mix by tapping.
   ※ In case of blood sample, Heparin, EDTA or citrate-treated whole blood are all suitable for this kit.
- 3) Transfer the lysate **into the Sample Well** of this kit, and wait until the sample solution is completely absorbed into the membrane. \*\* The amount of loading on the sample well should not exceeds 60 µl.
- 4) Go to  $\lceil$ 2. Common step $\rfloor$ .



## [Animal tissue, plant leaves or fruit]

- 1) Prepare 100 µl of Lysis Buffer into a 1.5 ml microcentrifuge tube (not provided).
- 2) Take a small piece of tissue (about ~5 mm diameter size) from animal or plant tissue.
- 3) Add tissue sample into the Lysis Buffer aliquot and crush the sample with a 100  $\mu$ l pipette tip by pressing it briefly against the tube wall.
  - \* If larger amount of tissue sample is used (>5mm), increase the volume of the lysis buffer to 200 μl.
- 4) Incubate at RT for 2 min for tissue lysis and nucleic acid releasing.
- 5) Spin briefly and transfer 50  $\mu$ l of the supernatant of lysate into the Sample Well of this kit, and wait until the sample solution is completely absorbed into the membrane.
- 6) Go to  $\lceil$ 2. Common step $\rfloor$ .



### [Plant seeds]

- 1) Plant seeds crush finely using a hammer, bead beater or tissuelyser.
- 2) Transfer crushed seed (about 10~20 mg) to 1.5 ml microcentrifuge tube (not provided).
- 3) Add 300 μl of Lysis Buffer and briefly mix by vortexing for 15 second.
   ※ If larger amount of seeds is used (>20mg), increase the volume of the lysis buffer to 500 μl.
- 3) Incubate at RT for 2 min for sample lysis and nucleic acid releasing.
- 4) Spin briefly and transfer 50  $\mu$ l of the supernatant of lysate into the Sample Well of this kit, and wait until the sample solution is completely absorbed into the membrane.
- 5) Go to  $\lceil$ 2. Common step $\rfloor$ .

# [Buccal swab]

- 1) Prepare **300** µl of Lysis Buffer into a 1.5 ml microcentrifuge tube (not provided).
- 2) Cut the cotton tip where the cells were collected.
- 3) Add cotton tip into the lysis buffer aliquot and briefly mix by vortexing or tapping.
- 4) Spin briefly and transfer 50  $\mu$ l of the supernatant into the Sample Well of this kit, and wait until the sample solution is completely absorbed into the membrane.
- 5) Go to  $\lceil$ 2. Common step $\rfloor$ .

#### [Urine]

- 1) Prepare 1 ml of sample into a 1.5 ml microcentrifuge tube (not provided), and centrifuge at 12,000 rpm for 1 min.
- 2) Remove supernatant by pipetting.
- 3) Add 50 µl of Lysis Buffer and mix by vortexing vigorously until the pellet resuspended completely.
- 4) Transfer lysate **into the Sample Well** of this kit, and wait until the sample solution is completely absorbed into the membrane.
- 5) Go to  $\lceil$ 2. Common step $\rfloor$ .



## [Soil or stool]

- 1) Prepare 10~20 mg of sample into a 1.5 ml microcentrifuge tube (not provided).
- 2) Add **500** µl of Lysis Buffer and mix by vortexing vigorously for 15 second.

  \*\* If larger amount of sample is used (>20 mg), increase the volume of the lysis buffer to 1 ml.
- 3) Centrifuge at 12,000 rpm for 1 min.
- 4) Transfer **50 μl of the supernatant into the Sample Well** of this kit and wait until the sample solution is completely absorbed into the membrane.
- 5) Go to  $\lceil$ 2. Common step $\rfloor$ .

### 2. Common step (After loading lysate to Sample Well)

- 1) Load 200 µl of Washing Buffer (80% ethanol) into the Washing Well and stay for 3~5 min.
  - **X** If the sample well is not clear after treatment of washing buffer, repeat this step with additional loading of 100 μl of Washing buffer.
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- 2) **Punch out about 1 mm size of membrane from the bottom of Sample Well** using a puncher or tip, and directly apply in PCR or RT-PCR reactions as template.
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  - **※** The nucleic acids on the kits are stable for 6 months at 4℃.

#### <Recommendation for PCR or RT-PCR>

- a) Optimal reaction volume for PCR or RT-PCR is 50 µl.
- b) Optimal cycle number is 35 ~ 45 cycles.
- c) Do not use high-fidelity PCR enzymes such as *Pfu* DNA polymerase or their derivatives.

#### **Products**

Cat. No.	Products	Size
PINA15	Punch-it™ NA-Sample Kit	15 Preps
PINA50	Punch-it™ NA-Sample Kit	50 Preps
PINA200	Punch-it™ NA-Sample Kit	200 Preps

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